



Single-cell RNA sequencing reveals intratumoral heterogeneity in primary uveal melanomas and identifies HES6 as a driver of the metastatic disease

Charlotte Pandiani^{1,2} · Thomas Strub^{1,2} · Nicolas Nottet^{1,2} · Yann Cheli^{1,2} · Giovanni Gambi³ · Karine Bille^{1,2} · Chrystel Husser^{1,2} · Mélanie Dalmasso^{1,2} · Guillaume Béranger^{1,2} · Sandra Lassalle^{1,4} · Virginie Magnone^{1,5} · Florence Pédeutour^{1,6} · Marie Irondelle^{1,7} · Célia Maschi^{1,8} · Sacha Nahon-Estève^{1,8} · Arnaud Martel^{1,8} · Jean-Pierre Caujolle^{1,8} · Paul Hofman^{1,4} · Kévin LeBrigand^{1,5} · Irwin Davidson³ · Stéphanie Baillif^{1,8} · Pascal Barbry^{1,5} · Robert Ballotti^{1,2} · Corine Bertolotto^{1,2}

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Abstract

Intratumor heterogeneity has been recognized in numerous cancers as a major source of metastatic dissemination. In uveal melanomas, the existence and identification of specific subpopulations, their biological function and their contribution to metastasis remain unknown. Here, in multiscale analyses using single-cell RNA sequencing of six different primary uveal melanomas, we uncover an intratumoral heterogeneity at the genomic and transcriptomic level. We identify distinct transcriptional cell states and diverse tumor-associated populations in a subset of the samples. We also decipher a gene regulatory network underlying an invasive and poor prognosis state driven in part by the transcription factor HES6. HES6 heterogenous expression has been validated by RNAscope assays within primary human uveal melanomas, which further unveils the existence of these cells conveying a dismal prognosis in tumors diagnosed with a favorable outcome using bulk analyses. Depletion of HES6 impairs proliferation, migration and metastatic dissemination in vitro and in vivo using the chick chorioallantoic membrane assay, demonstrating the essential role of HES6 in uveal melanomas. Thus, single-cell analysis offers an unprecedented view of primary uveal melanoma heterogeneity, identifies bona fide biomarkers for metastatic cells in the primary tumor, and reveals targetable modules driving growth and metastasis formation. Significantly, our findings demonstrate that HES6 is a valid target to stop uveal melanoma progression.

The authors contributed equally: Charlotte Pandiani, Thomas Strub, Robert Ballotti, Corine Bertolotto

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✉ Corine Bertolotto
corine.bertolotto-ballotti@inserm.fr

¹ Université Côte d'Azur, Nice, France

² Inserm, Biology and Pathologies of melanocytes, team1, Equipe labellisée Ligue 2020 and Equipe labellisée ARC 2019, Centre Méditerranéen de Médecine Moléculaire, Nice, France

³ Department of Functional Genomics and Cancer, Institut de Génétique et de Biologie Moléculaire et Cellulaire (IGBMC), Illkirch, France

⁴ Laboratoire de Pathologie clinique et expérimentale, biobanque BB-0033-00025, and IRCAN team 4, FHU OncoAge,

Introduction

Uveal melanoma is an aggressive and deadly neoplasm, which develops from melanocytes in the choroid. At diagnosis, only 1–3% of the patients have detectable metastases. Rapid local treatments are crucial, as survival correlates

Nice, France

⁵ CNRS, Institut de Pharmacologie Moléculaire et Cellulaire (IPMC), Valbonne, France

⁶ Laboratoire de Génétique des tumeurs solides and IRCAN, Nice, France

⁷ Inserm, Imagery platform, Centre Méditerranéen de Médecine Moléculaire, Nice, France

⁸ Pasteur 2 Teaching Hospital, Department of Ophthalmology, Nice, France

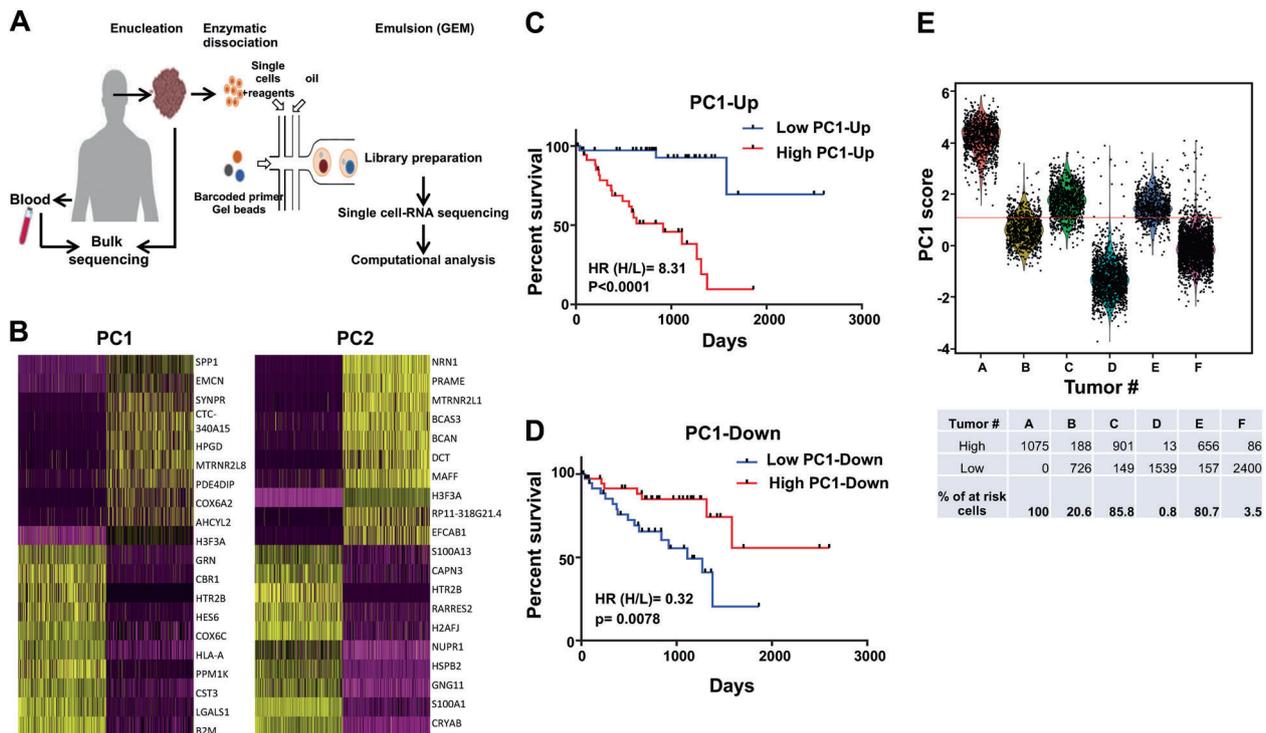


Fig. 1 Single-cell RNA-seq uncovers poor prognosis cell subpopulations. **a** Experimental workflow indicating the dissociation and isolation of individual cells from primary uveal melanomas for generating single-cell RNA-seq profiles. cDNA from the individual cells were synthesized, followed by library construction and massively parallel sequencing using the 10x genomic approach. **b** Heatmaps of the first two principal components from the principal component analysis (PCA) based on highly variable genes in the dataset. Both cells and genes are sorted by their PC scores allowing easy exploration of the principal sources of heterogeneity in the dataset. The first ten

genes with the highest or lowest absolute PC scores are shown for PC1 and PC2 (yellow). **c** Kaplan–Meier survival plot of the ten genes with the highest PC1 values (lower left quadrant of **b**). **d** Kaplan–Meier survival plot of ten genes with the lowest PC1 values (upper right quadrant of **b**). **e** Histograms showing PC1 score, based on the top ten up and down PC1 genes, of all the single cells in each tumor. The Youden index was used as the cutoff point (red line). The percentage of cells with high and low PC1 score as well as the percentage of “poor prognosis” cells per tumors are indicated below the histogram.

31 with primary tumor size [1]. Despite successful treatment of
 32 the primary tumor, metastases, that display a pronounced
 33 liver tropism, develop in 50% of patients within a median
 34 time of 2.4 years [2]. Once it has spread, there are no
 35 approved systemic treatments for uveal melanoma. Overall,
 36 90% of patients will die within 6 months after diagnosis of
 37 **Q6** metastases (reviewed in [3, 4]).

38 The above observations imply that cell subpopulations
 39 responsible for metastases, and patient death, disseminates
 40 early from the primary tumor. The identity of these cell
 41 subpopulations and the identification of their specific markers
 42 are required to improve patient outcome. Supporting
 43 this idea, in skin melanomas, intratumoral heterogeneity has
 44 been shown to have a profound impact on tumor evolution,
 45 development of metastases and therapy resistance [5–7].

46 Previous studies have separated uveal melanoma into
 47 two classes according to their transcriptomic profile. Class 1
 48 is predictive of poor metastatic risk and long-term survival,
 49 while class 2 is associated with a high risk of distant
 50 metastasis and a dismal prognosis [8]. However, to date
 51 none of the genes identified in these studies has been shown

to be potential therapeutic targets for uveal melanoma
 52 treatment.

53 Very recently, single-cell RNA-seq analyses provided a
 54 glimpse into primary and metastatic uveal melanomas
 55 ecosystems, and disclosed a regulatory T-cell phenotype,
 56 highlighting LAG3 as a potential candidate for immune
 57 checkpoint blockade [9]. LAG3 has also been pointed out to
 58 be a potential regulator of uveal melanoma immunity in
 59 other studies [10–12].

60 Thus, assessing intratumoral heterogeneity and char-
 61 acterization of the different transcriptional states, might
 62 provide insights into the subpopulation of uveal melanoma
 63 cells, that favor the metastatic dissemination and may lead
 64 to the identification of biomarkers to prevent the metastatic
 65 disease.
 66

Table 1 Histopathological, cytogenetic and genotypic features and classification for the six melanomas.

Tumor ID	LH16.3814	LH17.364	LH17.530	LH17.3222	LH17.3554	LH18.277
Tumor #	A	B	C	D	E	F
Sex	F	M	M	M	M	M
Age (year)	84	69	84	65	85	31
Largest basal diameter (mm)	14	18	19	10	15	17
Cell type	Spindle	Epithelioid	Mixed	Spindle	Spindle	Spindle
Mutation	GNAQ ^{Q209P}	GNA11 ^{Q209L}	WT	GNA11 ^{Q209L}	GNA11 ^{Q209L}	GNA11 ^{Q209L} , SF3B1 ^{R625H}
Chromosomal 8 Gain	K8q11.1-q24.3		K8q11.1-q24.3	K8q13.3-q24.3		K8q11.22-q24.3
Chromosomal 3 loss	K3		K3p12.3-p11.1; K3q13.11-q29		K3	
BAP1	Missense (c.91A)	Stopgain (c.829T)	Intronic (rs123602)	Intronic (rs419604; rs123602; rs409803)		WT
AJCC classification	pT3aNx	pT4aNx	pT4aNx	pT2aNx	pT2aNx	pT3bNx
Cytogenetic classification	2c	2a	2c	1b	2c	1b

Classification based on Trolet et al. [44].

AJCC American Joint Committee on Cancer, K chromosome.

67 Methods

68 Sample collection and processing

69 Single cells were isolated from tumor tissues (#A–F) of
70 patients diagnosed with ocular melanoma, after written
71 informed consent was obtained from the Nice CHU hospi-
72 tal. Samples were analyzed using the 10x Genomics's
73 protocol.

74 Cell cultures

75 Human uveal melanoma cell lines Mel270 (GNAQ^{Q209P})
76 [13], 92.1 (GNAQ^{Q209L}) [14], OMM2.5 (GNAQ^{Q209P}) [13]
77 and OMM1 (GNA11^{Q209L}) [15] were grown as previously
78 described. They all express BAP1. Additional information
79 about these cell lines may be found here [16]. MP46
80 (GNAQ^{Q209L}; no BAP1 expression) and MP65
81 (GNA11^{Q209L}; BAP1 c1717del) cell lines were from ATCC.
82 Cell lines are regularly tested for mycoplasma and are
83 mycoplasma-free.

84 RNAscope

85 mRNAs for *HES6* in sections from human uveal melanomas
86 were detected with RNAscope assay (Advanced Cell
87 Diagnostics, ACD) according to the manufacturer's proto-
88 cols. Images were captured with a confocal (Leica
89 DMI6000) microscope.

Statistical analyses

No statistical methods were used to determine sample size.
Sample size was determined to be adequate based on the
magnitude and consistency of measurable differences
between groups. The data are presented as the means ± SD
and analyzed using two-sided Student's *t*-test with Prism or
Microsoft Excel software (**p* value ≤ 0.05; ***p* value ≤ 0.01;
****p* value ≤ 0.001). For chick embryo chorioallantoic
membrane (CAM) assay, a one-way ANOVA analysis with
post-tests was done on the data.

Primers and siRNAs used

HES6 forward, TGA CCA CAG CCC AAA TTG C;
reverse, CTA CCC CAC CAC ATC TGA AC; *RPLP0*
forward, AAG GTG TAA TCC GTC TCC ACA GA;
reverse, CTA CCC CAC CAC ATC TGA AC. siRNA were
obtained from Sigma (#EHU036431) and Horizon Dis-
covery (#L-008408-00-0005).

Results

A subset of primary uveal melanomas displays intratumoral heterogeneity

To inspect intratumoral heterogeneity, we isolated indivi-
dual cells from six freshly resected human primary uveal
melanomas (#A–F) and generated single-cell transcriptomes
using 10x genomics (Fig. 1a). The clinical, histopathologic
and cytogenetic features are presented (Table 1).

Table 2 Cellular functions and diseases by second principal component (PC2).

Cellular movement	Cell movement of tumor cell lines	4.24E-18	3.061
Cellular movement	Migration of tumor cell lines	5.85E-16	3.009
Cell death and survival	Cell viability	2.80E-15	2.965
Cell death and Survival	Cell survival	3.47E-16	2.899
Cancer, organismal injury and abnormalities	Neoplasia of cells	1.89E-07	2.834
Cancer, gastrointestinal disease, hepatic system disease, organismal injury and abnormalities	Liver tumor	2.93E-11	2.695
Cell death and survival	Necrosis	2.89E-25	-2.085
Cell death and survival	Apoptosis	6.05E-25	-2.121
Cell death and survival	Cell death of connective tissue cells	2.14E-12	-2.276

Ingenuity pathway analysis (IPA) on the PC1 genes (z score $-1/+1$; 258 genes up; 15 down; minimum 30 molecules per modules).

115 Histological examination showed epithelioid cells in tumors
 116 B and C (Supplementary Fig. 1), and a high mitotic index in
 117 tumor B (not shown). Examination of copy number varia-
 118 tions (CNVs) by array comparative genomic hybridization
 119 (a-CGH) showed complete or partial loss of chromosome 3
 120 in tumors A, C, E and partial gain of chromosome 8 in
 121 tumors A, C, D and F (Supplementary Fig. 2). These CNVs
 122 are associated with high metastatic risk [17]. Finally, whole
 123 exome sequencing of bulk lesions indicated that all, except
 124 tumor C, carried activating mutation in GNAQ or GNA11,
 125 two frequent driver mutations in uveal melanomas
 126 (Table 1). Tumor C neither harbors mutations in *CYSLTR2*
 127 or *PLCB4* which mutations are also considered as driver
 128 events in uveal melanomas [18–20].

129 We first used the principal component analysis of which
 130 the two first principal components (PC) constituted the
 131 majority of the variance within the dataset (Fig. 1b and
 132 Supplementary Table 1). Among the ten genes with the
 133 highest PC1 and PC2 values was *HTR2B*, a gene previously
 134 associated with high metastatic risk and poor overall sur-
 135 vival [8]. Cellular function or disease analysis using Inge-
 136 nuity® Pathway Analysis (IPA) indicated that the
 137 PC1 signature (z score $-1/+1$; 258 genes up; 15 genes
 138 down) correlated with cell movement of tumor cell lines,
 139 migration of tumor cell lines, cell viability, cell survival,
 140 neoplasia of cells (Table 2). Interestingly, liver tumor
 141 function was also predicted and is consistent with a strong
 142 liver tropism of uveal melanomas. Conversely, cellular
 143 functions or diseases related to apoptosis or necrosis were
 144 inhibited. Analysis of upstream regulators with IPA high-
 145 lighted activation of transcription regulators and cytokines,
 146 with role in inflammation, and cellular stress, including
 147 STAT, NFKB, ATF6, XBP1, HIF1 and TNF proteins
 148 (Supplementary Table 2). IPA revealed that PC2 was also
 149 linked to proliferation of tumor cells and invasion of tumor
 150 (Supplementary Fig. 3a).

151 Kaplan–Meier analysis of uveal melanoma patients
 (TCGA set) showed that expression of the top ten genes

with the highest PC1 values was associated with shortened
 survival (Fig. 1c), whereas expression of the top ten genes
 with the lowest PC1 values correlated with an increased
 survival (Fig. 1d). Expression of the top ten genes with the
 highest values in PC2 was also predictive of a poor prog-
 nosis but that of the top ten genes with the lowest PC2
 values did not correlate with survival (Supplementary
 Fig. S3b, c).

To gain insights into the prognosis sensitivity and spe-
 cificity of the PC1 signature, we used the top ten up and
 down genes to calculate a “PC1 score” for each patient of
 the uveal melanoma TCGA cohort and plotted a ROC curve
 (Supplementary Fig. 4). The AUROC was 0.84 and the
 Youden index 0.63, thereby indicating that this “PC1 score”
 might be of interest to estimate patients’ prognosis. If we
 extrapolate this “score” to our single-cell analysis, cells with
 a “score” above the Youden index should be of “poor
 prognosis” i.e., expressed a gene signature associated with
 poor patient survival, while those with a “score” under the
 Youden index should be of “good prognosis”, because they
 expressed genes associated with low metastatic risk and
 long-term survival.

Applying this concept, we found that tumors A, C and E
 classified in the poor prognosis class 2c group by the
 cytogenetic analysis (Table 1), contained between 80 and
 100% of “poor prognosis cells”, while tumor B (class 2a,
 Table 1) contained only 20% (Fig. 1e). Among specimens
 with favorable predictable outcome (class 1b, Table 1),
 tumors D and F comprised only 0.8% and 3.5% of poor
 prognosis cells, respectively. This analysis, based on the
 “PC1 score” which reflects the ability of cells to metastasize
 and cause patient death, demonstrated an intratumoral
 transcriptomic and functional heterogeneity in uveal mel-
 anomas. On a clinical point of view, even a small number of
 cells with a high PC1 score might be sufficient to support
 distant metastasis development and impair patient survival.

To identify salient biological cell states, we next per-
 formed clustering of the individual cells with the Seurat

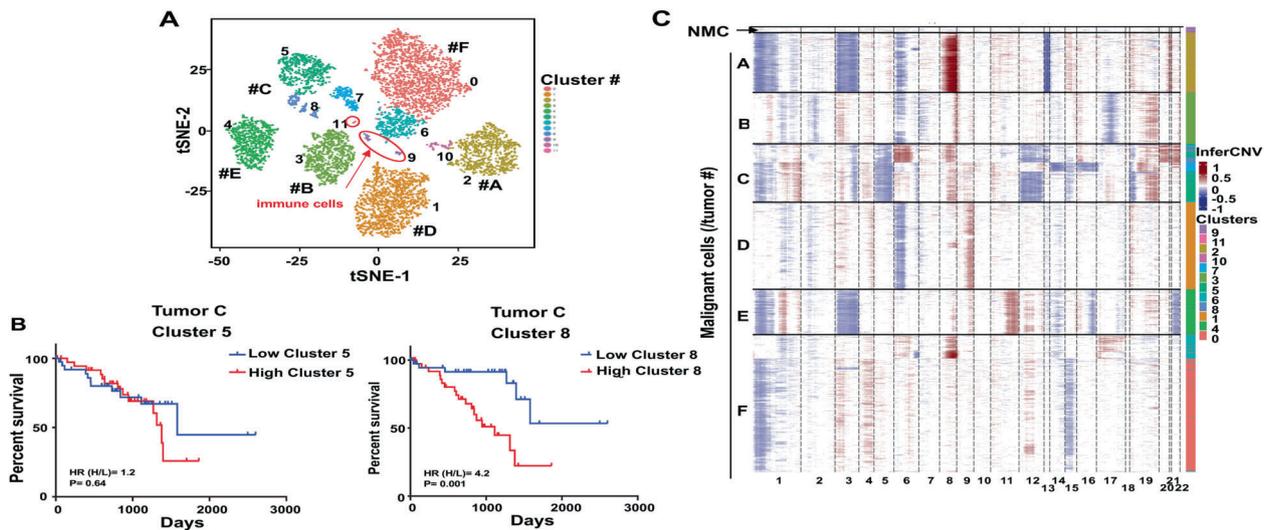


Fig. 2 Single-cell RNA-seq uncovers intratumoral heterogeneity. **a** Seurat analysis showing t-SNE plots of 7890 uveal melanoma cells colored by clusters. Each point represents a single cell. Red circles indicate non-malignant cells. **b** Kaplan–Meier survival plot for the top 20 genes of the indicated clusters. **c** Heatmap of inferred copy number variation (CNV) signal normalized against the topmost cluster

composed by the pool of all putative non-malignant cells (no CNV variation). Cells (rows, $n = 7890$ cells) are ordered from non-malignant cells (NMC, $n = 101$ cells) to cancer cells ($n = 7789$ cells), from the six uveal melanomas. Chromosomal regions (columns) with amplifications (red) or deletions (blue) are shown. The additional tracks, on the right, show the associated cluster number from Seurat.

191 analysis pipeline and used non-linear dimensionality
 192 reduction method [t-distributed stochastic neighbor
 193 embedding (t-SNE)], to visualize cell clusters. This analysis
 194 revealed that most cells grouped by tumor of origin, thereby
 195 indicating intertumor heterogeneity (Supplementary
 196 Fig. 5a). Further unbiased clustering of the individual cells
 197 identified 12 clusters (Fig. 2a). Tumors B, D and E each
 198 comprised a single cluster, while two clusters were identified
 199 in tumors A and F, and three clusters in tumor C, again
 200 emphasizing the existence of intratumor heterogeneity. Few
 201 non-malignant cells were detected in the tumors. Cluster 9
 202 was annotated as immune cells since it was enriched in the
 203 expression of T-cells and monocytes/macrophages markers
 204 and cluster 11 as endothelial cells since it was enriched in
 205 the expression of *PECAMI1*, *CD34*, *FLT1* *CDH5* (Supple-
 206 mentary Fig. 5b–d). These two latter clusters gathered by
 207 cell type and not by patient. Finally, representation of the
 208 cluster composition of each tumor, further demonstrated the
 209 transcriptomic heterogeneity of uveal melanoma cells
 210 within a tumor, and different cluster ratios in distinct tumors
 211 (Supplementary Fig. 6).

212 List of genes associated with each cluster (Supplemen-
 213 tary Table 3) was used in IPA comparison analysis to
 214 address enrichment in canonical pathways. Clusters 2, 4, 7,
 215 8 and 10 clustered together and disclosed clear activation of
 216 Rho GTPase-dependent signaling pathways, regulation of
 217 actin cytoskeleton and integrin signaling (Supplementary
 218 Fig. 7a). Equally important, in these clusters, Rho-GDI, a
 219 negative regulator of signaling through Rho GTPases, was
 220 downregulated. Other pathway more robustly expressed in

221 these clusters included mitochondria oxidative phosphor-
 222ylation [21]. In keeping with the recognized role of Rho
 223 GTPases and mitochondrial metabolism as markers of
 224 tumor invasion and metastasis, Kaplan–Meier survival plot
 225 generated from the top 20 genes in each cluster revealed
 226 that only clusters 2, 4, 7, 8 and 10 were associated with a
 227 poor prognosis (Fig. 2b and Supplementary Fig. 7b). In
 228 tumor C, whereas cluster 5 was not related to the prognosis,
 229 clusters 7 and 8 contained cells conveying a dismal prog-
 230 nosis, further supporting the existence of transcriptomic and
 231 functional intratumoral heterogeneity in primary uveal
 232 melanomas (Fig. 2b and Supplementary Fig. 7b).

233 In addition, as previously described [22], large-scale
 234 copy number aberrations for each cell by averaging relative
 235 expression levels over large genomic regions was used to
 236 infer CNVs from scRNA-seq data (Fig. 2c). Inferred-CNV
 237 profiles uncovered distinct chromosomal imbalance,
 238 including chromosome 3 loss and 8q gain, that are char-
 239 acteristic uveal melanoma alterations. However, tumors C
 240 and F appeared to contain more than one genetic clone.
 241 Cryptic alterations, in cell subsets of tumors A and D can
 242 also be observed in chromosomes 6 and 8, respectively.
 243 Globally, inferred-CNV analysis was in agreement with
 244 bulk array-based CGH (Supplementary Fig. 2).

245 Collectively, in addition to intertumor heterogeneity, this
 246 dataset discloses an intratumoral heterogeneity at both the
 247 genetic and transcriptomic level in a subset of tumors.
 248 Transcriptomic and genetic heterogeneity overlapped lar-
 249 gely in tumor F where the cells with 8q gain fell in cluster 6,
 250 and in tumor C, where cells having a loss in chromosomes

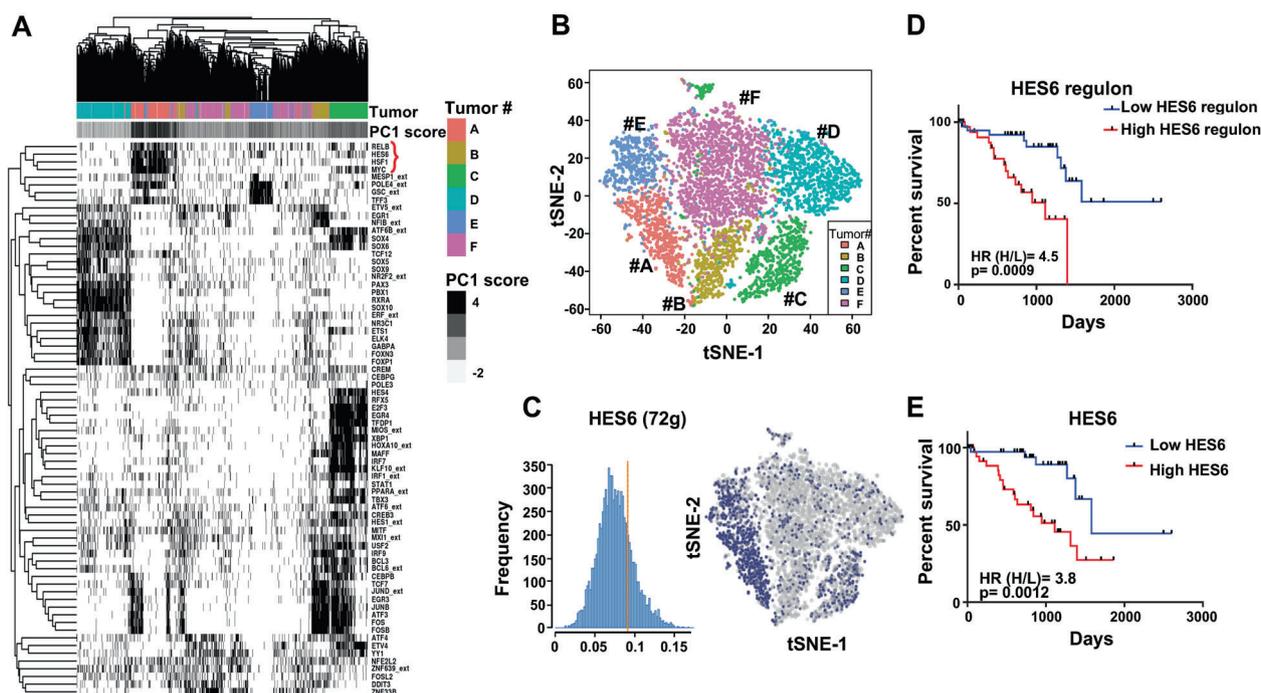


Fig. 3 scRNA-Seq identifies multiple co-existing transcriptional states and a network driven by HES6 associated with a poor prognosis. **a** Heatmap of cells and regulon binary scores with hierarchical clustering inferred by SCENIC. The additional track, above, show supervised clustering by patients and the PC1 score based on the top ten genes with the highest values in PC1. The 72 regulons with the best clustering out of the 122 identified in the six primary uveal melanomas are represented. **b** t-SNE shows cells colored by patient

(SCENIC approach). **c** HES6 regulon activity comprising 72 genes quantified using AUCell is represented. Regulons of predicted transcription factors in the six patients were determined to be active if they exceeded the threshold (Blue), otherwise, regulons were considered to be inactive (gray). **d** Kaplan–Meier survival plot of the HES6 regulon (TCGA dataset). **e** Kaplan–Meier survival plot of HES6 (TCGA dataset).

14, 15 and 16 segregated to cluster 7. However, in tumor C, cells with a 6p gain were distributed between cluster 8 and a portion of cluster 5. Importantly, minority pre-existing subclones or rare poor prognosis cells may be missed by classical bulk sequencing because their abundance falls below the lower limit of sensitivity, while they represent the functional cell subsets that will outgrowth and drive the metastatic dissemination.

Characterization of major cell subpopulations in primary uveal melanomas

Next, to get insights into the stable transcriptional cell states, we used the Single-cell regulatory network inference and clustering (SCENIC) method [23]. SCENIC exploits transcription factors and cis-regulatory sequences, to map the activity of the regulatory networks (regulons) underlying the different gene expression signatures. This analysis disclosed 122 regulons (out of 1046) that displayed significant activity in uveal melanomas (Fig. 3a and Supplementary Fig. 8). After non-linear dimensionality reduction (t-SNE) of these data, we can observe a degree of cellular overlapping between cells from different tumors, indicating that cells with similar transcriptional program can be found

in different tumors (Fig. 3b). Together, these findings further confirm the transcriptional intratumor heterogeneity.

SCENIC heatmap also revealed clustered regulons (RELB, HES6, HSF1 and MYC) that correlated with a high PC1 score. This transcriptional state can be inferred as an invasive state as *MYC* and *RELB* have been involved in metastasis of uveal melanoma cells [24–28]. However, we focused our attention on HES6 (enhancer of split family basic helix-loop-helix transcription factor 6). HES6 was detected among the top ten genes with the highest PC1 values, it stimulates the invasive ability of various tumor cells [25–27], and its role in uveal melanomas remains to be elucidated. Cells with high HES6 regulon activity were found mainly in tumors A, C and E, but few HES6-positive cells can be found in other tumors (Fig. 3c).

Importantly, Kaplan–Meier analysis showed that the *HES6* regulon (Supplementary Table 4) as well *HES6* itself, which is carried by chromosome 2, were negatively correlated with overall survival (Fig. 3d, e). In keeping with this, both in tumors analyzed hereby and the TCGA dataset, *HES6* expression is associated with chromosome 3 loss (Supplementary Fig. 9), which in uveal melanomas highly correlates with the metastatic risk. Notably, *HES6* expression overlapped with class 1b and class 2 tumors [8], which

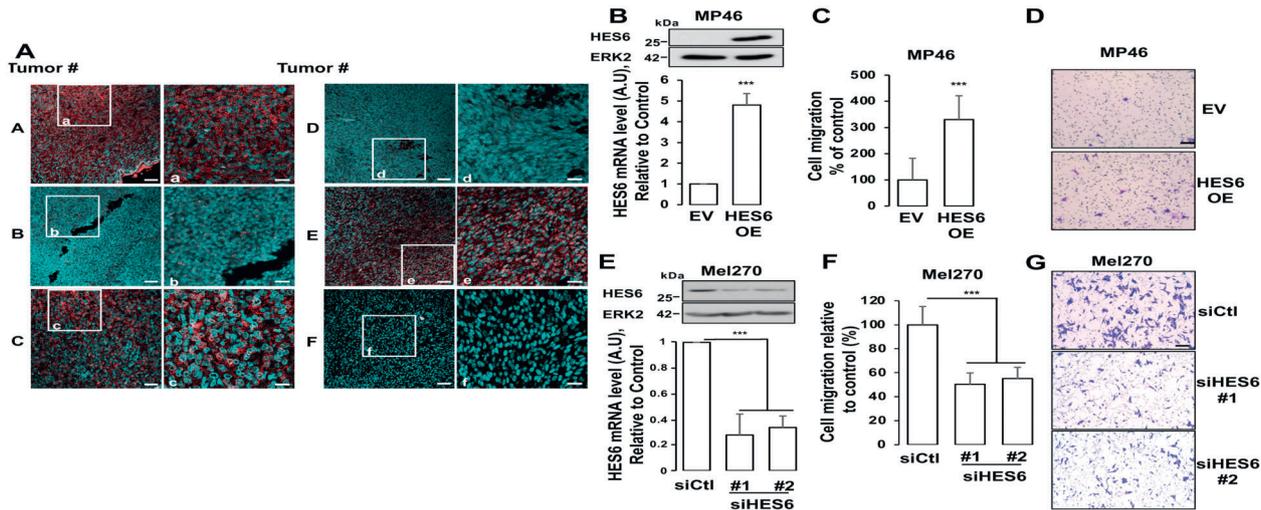


Fig. 4 HES6 expression controls the motile ability of primary uveal melanomas. **a** Sections from tumors # A–F were labeled with RNAscope probe for *HES6* (red), and images were captured by confocal microscopy. Cell nuclei (blue green). Shown are the areas of high and low heterogeneity. Scale bars represent 60 μ m. For each tumor, magnification of the boxed area is shown (Right). Scale bars represent 25 μ m. **b** Western blot and RT-QPCR analysis of *HES6* in primary MP46 melanoma cells transduced with a control or HES6 adenovirus expressing HES6 (HES6 OE) for 72 h. **c** Human primary MP46 melanoma cells were transduced with empty (control) or HES6 expressing adenovirus

(HES6 OE) for 48 h before being seeded in the upper part of the Boyden chamber. Migration was examined after 24 h. Values represent means +SD of three independent experiments. $^{**}p < 0.01$. **d** Representative images are shown. Bar = 100 μ m. **e** Western blot and RT-QPCR analysis of *HES6* in primary Mel270 melanoma cells transfected with a control siRNA (siCtl) or two different pools of multiple siRNA targeting HES6 (siHES6#1 and siHES6#2). **f** Migration of Mel270 cells transfected with the indicated siRNA. $^{***}p < 0.001$. **g** Representative images are shown. Bar = 100 μ m.

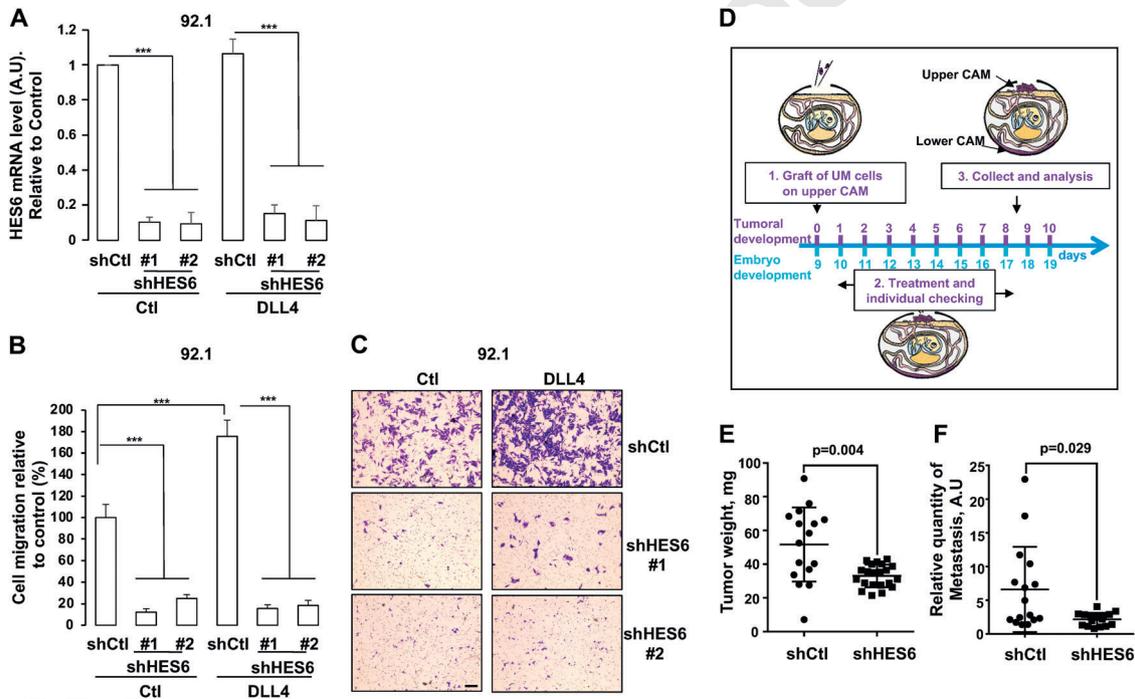


Fig. 5 HES6 signaling is a key driver of aggressive and motile phenotypes. **a** QPCR analysis of *HES6* in primary 92.1 melanoma cells expressing doxycycline-inducible control or HES6 shRNA in presence of 1 μ g/ml doxycycline for 96 h in presence or absence of DLL4 1 μ g/ml. **b** Migration of 92.1 melanoma cells expressing doxycycline-inducible control or HES6 shRNA in presence of 1 μ g/ml doxycycline for 48 h in presence or absence of DLL4 1 μ g/ml. $^{***}p < 0.001$. **c** Representative images are shown. Bar = 100 μ m. **d** Description of the chicken embryo

CAM assay. **e** 92.1 uveal melanoma cells expressing doxycycline-inducible control or HES6 shRNA were grafted on the CAM of 9-day-old (E9) chick embryos. The tumors were collected and weighted on day 18 (E18). Values represent means + SEM. **f** Genomic DNA is extracted from the lower CAM to evaluate the number of metastatic cells on day 18 and analyzed by qPCR with specific primers for human Alu sequences. Values represent means + SEM.

297 bear worse prognosis than class 1a (Supplementary Fig. 10).
 298 Taken together these observations highlight the role of
 299 HES6 as a key marker of uveal melanoma cell metastatic
 300 potential and patient survival.

301 **HES6 enhances growth and motile ability in vitro** 302 **and in vivo of primary uveal melanoma**

303 To validate these analyses and given the lack of high-
 304 quality HES6 antibody for immunohistochemistry, its expression
 305 in human patient biopsies was evaluated by RNAscope®
 306 fluorescence in situ hybridization assay. The staining con-
 307 firmed that primary uveal melanomas comprised both
 308 HES6-high and HES6-low cells that were segregated or
 309 intermixed reflecting regional heterogeneity and different
 310 cell states (Fig. 4a). Negative control staining is shown
 311 (Supplementary Fig. 11). In line with the single-cell anal-
 312 ysis, HES6 expression was higher in tumors A, C and E
 313 compared to tumors B, D and F.

314 Next, we aimed to portray the biological role of HES6.
 315 We first assessed the ability of HES6 to control the motile
 316 capacity of primary uveal melanoma cells. Ectopic HES6
 317 expression enhanced migration of two different primary cell
 318 lines (Fig. 4b–d and Supplementary Fig. 12a, c). Con-
 319 versely, reduced migration of primary cells was obtained
 320 with HES6 inhibition by both siRNA and shRNA
 321 (Figs. 4e–g and 5a–c). Although we searched for metastatic
 322 drivers in the primary tumor, we also asked whether HES6
 323 could have an effect in the metastatic settings. Our results
 324 showed that HES6 gain enhanced (Supplementary
 325 Fig. 12d–f), whereas HES6 loss reduced (Supplementary
 326 Fig. 13a, b) motility of metastatic cells.

327 HES6 inhibition by siRNA or by using an inducible
 328 shRNA strategy also prevented the ability to form colony in
 329 primary (Supplementary Fig. 14a–c). The same held true in
 330 metastatic cells (Supplementary Fig. 14d–f). Thus, our
 331 findings indicated that HES6 might represent a valid target
 332 to limit uveal melanoma cell proliferation and migration.

333 *HES6* is an atypical *HES* gene whose role as downstream
 334 effector of NOTCH signaling is unclear. Among NOTCH
 335 natural ligand, in uveal melanomas, Delta-like ligand 4
 336 (DLL4) is the NOTCH ligand the most associated with the
 337 metastatic risk and its expression is the most inversely
 338 correlated with patient survival (Supplementary Fig. 15a).
 339 Although a role for NOTCH signaling pathway has been
 340 reported in uveal melanoma [28, 29], the effect of DLL4 has
 341 never been investigated. Our data showed that DLL4
 342 increased NOTCH reporter activity, an effect that was
 343 inhibited by the γ -secretase inhibitor BMS-906024 (Sup-
 344 plementary Fig. 15b). In addition, blocking the NOTCH
 345 pathway with two NOTCH inhibitors BMS-906024 and
 346 DAPT reduced spheroid formation (Supplementary
 347 Fig. 15c). Finally, we observed that DLL4 enhanced uveal

melanoma cell migration (Supplementary Fig. 15d). Thus,
 DLL4 activates the NOTCH signaling pathway in uveal
 melanoma cells and controls their growth and migration.

To delineate the role of HES6 downstream of NOTCH, we
 assessed the impact of HES6 knockdown upon treatment with
 DLL4. Compared to control, DLL4 could no longer increase
 the migration of HES6 knockdown 92.1 and Mel270 cell lines
 (Fig. 5a–c and Supplementary Fig. 16a, b). These data pro-
 vide evidences that HES6 has critical tumorigenic properties
 downstream the NOTCH signaling pathway and mediates its
 effect on the motile ability of primary uveal melanoma cells.

Next, we demonstrated that HES6 knockdown in 92.1
 and Mel270 cells also reduced the formation of 3D spher-
 oids, that more faithfully model the tumor microenviron-
 ment than 2D cell cultures (Supplementary Fig. 17a–f).
 Further, a matrigel invasion assay showed that Mel270 cells
 originating from the control spheroids efficiently invaded
 the matrigel compared to spheroids formed with the HES6-
 knockdown cells (Supplementary Fig. 17g). This experi-
 ment could not be performed with 92.1 cells given that
 HES6 knockdown dramatically reduced sphere formation in
 these cells, thereby preventing spheroids for being har-
 vested and embedded in matrigel. Thus, HES6 also controls
 the invasive ability of primary uveal melanoma cells.

We reasoned that HES6 might be effective in driving
 metastatic dissemination of primary uveal melanoma cells.
 We thus studied tumor progression to metastasis of primary
 uveal melanoma cell in vivo using the CAM model
 (Fig. 5d). Control cells efficiently formed tumors and were
 overall also very efficient at forming metastasis as evi-
 denced by cells that had disseminated to the lower CAM
 (Fig. 5d–f). Growth and metastatic abilities were strongly
 reduced by 36% and 48%, respectively, with tumors formed
 from HES6 knocked-down cells.

Altogether, our findings showed both in vitro and in vivo
 that HES6 stimulates the aggressive potential of primary
 uveal melanoma and their motile capacity.

335 **Discussion**

336 Here, we used a single-cell transcriptomic profiling strategy
 337 to address the critical questions of cell heterogeneity in
 338 primary uveal melanomas in order to identify cell sub-
 339 populations driving the metastatic process.

340 The data gathered hereby, while confirming the existence
 341 of an intertumor heterogeneity, also uncover a molecular
 342 and functional intratumor heterogeneity. They highlight a
 343 new signature that allows to detect tumor cells that might
 344 convey unfavorable outcome among patients classified as
 345 having a good prognosis either by using classical clinical
 346 parameters or even gene expression profile on bulk tumor.
 347 An intratumoral genomic heterogeneity has previously been

suspected, since DNA extracted from several areas within the same primary uveal melanomas displayed different chromosomal abnormalities [30]. Our study discloses a transcriptomic heterogeneity that is not always supported by the genomic heterogeneity, but that reflects different transcriptional programs.

SCENIC has inferred at least three transcriptional states. One is related to cell specification due to the enrichment in SOX9, SOX10 and PAX3 regulons. This state overlaps with cells displaying intermediate activity of MITF a master regulator of melanocyte differentiation, proliferation and survival [31, 32]. SOX10 and PAX3 activity was inferred by SCENIC in cells with low PC1 score (good prognosis). In line with this, uveal melanoma patients from the TCGA cohort with high levels of both PAX3 and SOX10 have an increased overall survival (not shown).

A second transcriptional cell state, with enrichment in regulons BCL3, CEBPB and AP1 members (*JUNB*, *JUND*, *FOS* and *FOSB*), may be related to immune response and inflammation. Indeed, BCL3 and CEBPB have direct roles in the regulation of proinflammatory cytokine production by cancer cells [33–36]. Further, in cutaneous melanomas, activation of JUN leads to melanoma cell dedifferentiation via MITF downregulation that is associated with the production of proinflammatory cytokines [37, 38]. This transcriptional profile defined a primary uveal melanoma intrinsic inflammatory state that should favor immune cell infiltrate. However, none of the tumors inspected in our study showed a significant immune cell infiltration. This observation is in agreement with previous work from the TCGA network also reporting immune infiltration in a few numbers of primary uveal melanomas [39].

Finally, we focused our attention on the third transcriptional state inferred by SCENIC with invasive functionality that is driven, at least in part, by HES6. HES6, that belongs to the poor prognosis signature we discovered (top ten genes of PC1), is an atypical HES gene whose role in uveal melanomas remained totally unknown. By contrast to canonical NOTCH targets, HES6 was thought to antagonize NOTCH signaling. However, in uveal melanoma cells, HES6 knockdown impairs migration induced by DLL4, an activator of NOTCH receptors, indicating that HES6, depending on the context, may be a NOTCH effector. We demonstrate in vitro and in vivo that HES6 is a key driver of uveal melanoma proliferation and metastatic dissemination. Our data reveal that the subgroup of regulons activated in cells with a high PC1 score and therefore with poor survival prognosis displayed in addition to the HES6 regulon, the MYC regulon and was also partially enriched in JUN (*JUNB* and *JUND*) regulons. Increased MYC and JUN activities were also pinpointed in poor prognosis class 2 tumors by a previous single-cell analysis [9]. The work of Durante et al. also identified the activation of ARNT, TAF1

and TAF7 regulons in poor prognosis cells that were not spotted in our study. Conversely, HES6 and HES6 regulon, that are clearly associated with decreased survival, were not identified by Durante et al. Differences with our study can be explained by the fact that Durante et al. analyzed a mix of primary and metastatic specimens while in our study, we only focused on primary melanomas [9]. Further, they selected tumor cells using the expression of the differentiation markers DCT, MITF and MelanA [9]. Depending on the threshold, this filtering may induce biases by missing some cells in the analyses. Finally, Durante et al. analyzed a subgroup of uveal melanomas with a large immune infiltrate that could potentially affect tumoral cells transcriptomic profile [9]. Nevertheless, the work from Durante et al. shows important data about uveal melanoma ecosystem [9]. By contrast, our study, focusing on primary uveal melanomas with low immune infiltrate, which represent the vast majority of uveal melanoma (TCGA), discloses new transcriptomic signatures and pathways that are associated with prognosis and have direct impact on the biology of uveal melanoma cells.

In keeping with a role of HES6 in invasive ability, IPA analysis of the PC1 signature reveals activation of the Rho GTPase and integrin signaling pathways in cell subpopulations that convey a poor prognosis. Rho GTPases are essential in propagating integrin-mediated responses and, by tightly regulating actin cytoskeleton, offer a key signaling link through which adhesion, spreading, and migration are controlled in tumor cells [40]. Further, Rho lies downstream from GNAQ and GNA11 and stimulates YAP, which in turn controls uveal melanoma cell proliferation [41]. Of note, PAX3 can use YAP as a coactivator. Mechanistically, YAP activation can lead to the stimulation of PAX3 driven differentiation program [42], while in absence of PAX3, YAP is made available for TEAD transcription factors to drive uveal melanoma cell proliferation. This might append an additional level of heterogeneity.

Likewise, enhanced HES6 expression stimulates the invasive phenotype of prostate cancer, glioma and colorectal cancer cells [25, 26]. Conversely, HES6 knockdown has been reported to decrease migration of glioma, glioblastoma, colorectal cancer cells and of alveolar rhabdomyosarcoma [27, 43].

Whether the cell states and key transcription factors identified in primary lesions are maintained in the subsequent metastasis and play a critical role remains to be verified. However, in favor of this idea, HES6 knockdown also reduced growth and motile ability of metastatic uveal melanoma cells.

The identification of a HES6-driven transcriptional state, which is associated with high tumorigenic properties, is highly relevant for patient care, since we demonstrated that tumors classified as of good prognosis using bulk analysis,

504 contained varying proportions of HES6-positive cells, that
505 could negatively impact on patient outcome.

506 Given the lack of treatment options for metastatic uveal
507 melanomas, HES6 or its target genes that we disclose
508 hereby may represent actionable factors to be targeted
509 therapeutically.

510 Thus, our single-cell transcriptomic profiling uncovers
511 the existence of intratumor heterogeneity in primary uveal
512 melanomas and leads to mechanistic insights into the reg-
513 ulation of the metastatic process in uveal melanomas,
514 thereby offering unprecedentedly described biomarkers
515 with critical implications for prognosis and therapeutic
516 strategies.

517 Data availability

518 The experimental data from single-cell RNA sequencing,
519 whole exome sequencing and array-CGH have been
520 deposited in the NCBI Gene Expression Omnibus (GEO)
521 database (<https://www.ncbi.nlm.nih.gov/geo/>) under the
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540 **Author contributions** CB and RB designed and supervised the study,
541 prepared the figures and wrote the manuscript. VM carried out scRNA-
542 seq experiments. KLB and NN performed the computational analyses
543 and assisted in data analysis in discussions with PB who also critically
544 reviewed the manuscript. CP, TS, YC and GB performed patient
545 sample processing and functional experiments. KB, CH, MD and MI
546 provided technical assistance. GG and ID carried out the RNAscope
547 assays. SL and PH coordinated patient sample collection, maintained
548 IRB approval and performed histological analysis. FP provided cyto-
549 genetic analyses. AM, SNE, CM, JPC and SB gathered patients’
550 consent, provided the samples and clinical data.

551 Compliance with ethical standards

552 **Ethics** The study was approved by the hospital ethics committee (Nice
553 Hospital Center and University of Nice Côte d’Azur). The study was
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References

1. Rietschel P, Panageas KS, Hanlon C, Patel A, Abramson DH, Chapman PB. Variates of survival in metastatic uveal melanoma. *J Clin Oncol.* 2005;23:8076–80. 561
2. Nabil AA, Marie S, Marc-Henri S, Nathalie C, Laurence D, Sophie PN, et al. Upcoming translational challenges for uveal melanoma. *Br J Cancer.* 2015;113:1249–53. 562
3. Pandiani C, Beranger GE, Leclerc J, Ballotti R, Bertolotto C. Focus on cutaneous and uveal melanoma specificities. *Genes Dev.* 2017;31:724–43. 563
4. Yang J, Manson DK, Marr BP, Carvajal RD. Treatment of uveal melanoma: where are we now? *Ther Adv Med Oncol.* 2018. <https://doi.org/10.1177/1758834018757175>. 564
5. Tirosh I, Izar B, Prakadan SM, Wadsworth MH 2nd, Treacy D, Trombetta JJ, et al. Dissecting the multicellular ecosystem of metastatic melanoma by single-cell RNA-seq. *Science.* 2016;352:189–96. 565
6. Kemper K, Krijgsman O, Cornelissen-Steijger P, Shahrabi A, Weeber F, Song JY, et al. Intra- and inter-tumor heterogeneity in a vemurafenib-resistant melanoma patient and derived xenografts. *EMBO Mol Med.* 2015;7:1104–18. 566
7. Rambow F, Marine J, Goding CR. Melanoma plasticity and phenotypic diversity: therapeutic barriers and opportunities. *Genes Dev.* 2019;33:1295–318. 567
8. Onken MD, Worley LA, Ehlers JP, Harbour JW. Gene expression profiling in uveal melanoma reveals two molecular classes and predicts metastatic death. *Cancer Res.* 2004;64:7205–9. 568
9. Durante MA, Rodriguez DA, Kurtenbach S, Kuznetsov JN, Sanchez MI, Decatur CL, et al. Single-cell analysis reveals new evolutionary complexity in uveal melanoma. *Nat Commun.* 2020;11:496. 569
10. Karlsson J, Nilsson LM, Mitra S, Alsén S, Shelke GV, Sah VR, et al. Molecular profiling of driver events in metastatic uveal melanoma. *Nat Commun.* 2020;11:1894. 570
11. Figueiredo CR, Kalirai H, Sacco JJ, Azevedo RA, Duckworth A, Slupsky JR, et al. Loss of BAP1 expression is associated with an immunosuppressive microenvironment in uveal melanoma, with implications for immunotherapy development. *J Pathol.* 2020;250:420–39. 571
12. Triozzi PL, Schoenfeld L, Plesec T, Sauntharajah Y, Tubbs RR, Singh AD. Molecular profiling of primary uveal melanomas with tumor-infiltrating lymphocytes. *Oncoimmunology.* 2014;8:e947169. 572
13. Chen PW, Murray TG, Uno T, Salgaller ML, Reddy RKB. Expression of MAGE genes in ocular melanoma during progression from primary to metastatic disease. *Clin Exp Metastasis.* 1997;15:509–18. 573
14. De Waard-Siebinga I, Blom DR, Griffioen M, Schrier PI, Hoogendoorn E, Beverstock G, et al. Establishment and characterization of an uveal-melanoma cell line. *Int J Cancer.* 1995;62:155–61. 574
15. Luyten GP, Naus NC, Mooy CM, Hagemeyer A, Kan-Mitchell J, Van Drunen E, et al. Establishment and characterization of primary and metastatic uveal melanoma cell lines. *Int J Cancer.* 1996;66:380–7. 575
16. Griewank KG, Yu X, Khalili J, Sozen MM, Stempke-Hale K, Bernatchez C, et al. Genetic and molecular characterization of 576

- 616 uveal melanoma cell lines. *Pigment Cell Melanoma Res.* 2012;25:182–7. 666
- 617 17. Shain AH, Bagger MM, Yu R, Chang D, Liu S, Vemula S, et al. 667
- 618 The genetic evolution of metastatic uveal melanoma. *Nat Genet.* 668
- 619 2019;51:1123–30. 669
- 620 18. Larribère L, Utikal J. Update on gna alterations in cancer: 670
- 621 Implications for uveal melanoma treatment. *Cancers.* 671
- 622 2020;12:1–18. 672
- 623 19. Johansson P, Aoude LG, Wadt K, Glasson WJ, Warriar SK, 673
- 624 Hewitt AW, et al. Deep sequencing of uveal melanoma identifies a 674
- 625 recurrent mutation in PLCB4. *Oncotarget.* 2016;7:4624–31. 675
- 626 20. Moore AR, Ceraudo E, Sher JJ, Guan Y, Shoushtari AN, Chang 676
- 627 MT, et al. Recurrent activating mutations of G-protein-coupled 677
- 628 receptor CYSLTR2 in uveal melanoma. *Nat Genet.* 678
- 629 2016;48:675–80. 679
- 630 21. Zhao J, Zhang J, Yu M, Xie Y, Huang Y, Wolff DW, et al. 680
- 631 Mitochondrial dynamics regulates migration and invasion of 681
- 632 breast cancer cells. *Oncogene.* 2013;32:4814–24. 682
- 633 22. Patel AP, Tirosh I, Trombetta JJ, Shalek AK, Gillespie SM, 683
- 634 Wakimoto H, et al. Single-cell RNA-seq highlights intratumoral 684
- 635 heterogeneity in primary glioblastoma. *Science.* 685
- 636 2014;344:1396–401. 686
- 637 23. Aibar S, Gonzalez-Blas CB, Moerman T, Huynh-Thu VA, Imri- 687
- 638 chova H, Hulselmans G, et al. SCENIC: single-cell regulatory 688
- 639 network inference and clustering. *Nat Methods.* 2017;14:1083–6. 689
- 640 24. Meir T, Dror R, Yu X, Qian J, Simon I, Pe'er J, et al. Molecular 690
- 641 characteristics of liver metastases from uveal melanoma. *Invest* 691
- 642 *Ophthalmol Vis Sci.* 2007;48:4890–6. 692
- 643 25. Xu Y, Liu X, Zhang H, Zhu Z, Wu X, Wu X, et al. Over- 693
- 644 expression of HES6 has prognostic value and promotes metastasis 694
- 645 via the Wnt/beta-catenin signaling pathway in colorectal cancer. 695
- 646 *Oncol Rep.* 2018;40:1261–74. 696
- 647 26. Haapa-Paananen S, Kiviluoto S, Waltari M, Puputti M, Mpindi JP, 697
- 648 Kohonen P, et al. HES6 gene is selectively overexpressed in 698
- 649 glioma and represents an important transcriptional regulator of 699
- 650 glioma proliferation. *Oncogene.* 2012;31:1299–310. 700
- 651 27. Wickramasinghe CM, Domaschitz R, Amagase Y, Williamson 701
- 652 D, Missiaglia E, Shipley J, et al. HES6 enhances the motility of 702
- 653 alveolar rhabdomyosarcoma cells. *Exp Cell Res.* 703
- 654 2013;319:103–12. 704
- 655 28. Asnagli L, Ebrahimi KB, Schreck KC, Bar EE, Coonfield ML, 705
- 656 Bell WR, et al. Notch signaling promotes growth and invasion in 706
- 657 uveal melanoma. *Clin Cancer Res.* 2012;18:654–65. 707
- 658 29. Asnagli L, Handa JT, Merbs SL, Harbour JW, Eberhart CG. A 708
- 659 role for Jag2 in promoting uveal melanoma dissemination and 709
- 660 growth. *Investig Ophthalmol Vis Sci.* 2013;54:295–306. 710
- 661 30. Dopierala J, Damato BE, Lake SL, Taktak AF, Coupland SE. 711
- 662 Genetic heterogeneity in uveal melanoma assessed by multiplex 712
- 663 ligation-dependent probe amplification. *Invest Ophthalmol Vis* 713
- 664 *Sci.* 2010;51:4898–905. 714
31. Goding CR, Arnheiter H. MITF—the first 25 years. *Genes Dev.* 666
- 2019;33:983–1007. 667
32. Cheli Y, Ohanna M, Ballotti R, Bertolotto C. Fifteen-year quest 668
- for microphthalmia-associated transcription factor target genes. 669
- Pigment Cell Melanoma Res.* 2010;23:27–40. 670
33. Ohanna M, Giuliano S, Bonet C, Imbert V, Hofman V, Zangari J, 671
- et al. Senescent cells develop a parp-1 and nuclear factor-κB- 672
- associated secretome (PNAS). *Genes Dev.* 2011;25:1245–61. 673
34. Ohanna M, Cheli Y, Bonet C, Bonazzi VF, Allegra M, Giuliano S, 674
- et al. Secretome from senescent melanoma engages the STAT3 675
- pathway to favor reprogramming of naive melanoma towards a 676
- tumor-initiating cell phenotype. *Oncotarget.* 2013;4:2212–24. 677
35. Kuilman T, Michaloglou C, Vredeveld LC, Douma S, van Doorn 678
- R, Desmet CJ, et al. Oncogene-induced senescence relayed by an 679
- interleukin-dependent inflammatory network. *Cell.* 680
- 2008;133:1019–31. 681
36. Chang TP, Vancurova I. Bcl3 regulates pro-survival and pro- 682
- inflammatory gene expression in cutaneous T-cell lymphoma. 683
- Biochim Biophys Acta - Mol Cell Res.* 2014;1843:2620–30. 684
37. Riesenbergs S, Groetchen A, Siddaway R, Bald T, Reinhardt J, 685
- Smorra D, et al. MITF and c-Jun antagonism interconnects mel- 686
- anoma dedifferentiation with pro-inflammatory cytokine respon- 687
- siveness and myeloid cell recruitment. *Nat Commun.* 688
- 2015;6:8755. 689
38. Landsberg J, Kohlmeyer J, Renn M, Bald T, Rogava M, Cron M, 690
- et al. Melanomas resist T-cell therapy through inflammation- 691
- induced reversible dedifferentiation. *Nature.* 2012;490:412–6. 692
39. Robertson AG, Shih J, Yau C, Gibb EA, Oba J, Mungall KL, et al. 693
- Integrative analysis identifies four molecular and clinical subsets 694
- in uveal melanoma. *Cancer Cell.* 2017;32:204–20 e15. 695
40. Lawson CD, BurrIDGE K. The on-off relationship of Rho and Rac 696
- during integrin-mediated adhesion and cell migration. *Small* 697
- GTPases.* 2014;5:e27958. 698
41. Feng X, Degese MS, Iglesias-Bartolome R, Vaque JP, Molinolo 699
- AA, Rodrigues M, et al. Hippo-independent activation of YAP by 700
- the GNAQ uveal melanoma oncogene through a trio-regulated rho 701
- GTPase signaling circuitry. *Cancer Cell.* 2014;25:831–45. 702
42. Miskolczi Z, Smith MP, Rowling EJ, Ferguson J, Barriuso J, 703
- Wellbrock C. Collagen abundance controls melanoma phenotypes 704
- through lineage-specific microenvironment sensing. *Oncogene.* 705
- 2018;37:3166–82. 706
43. Roma J, Masià A, Reventós J, De Toledo JS, Gallego S. Notch 707
- pathway inhibition significantly reduces rhabdomyosarcoma 708
- invasiveness and mobility in vitro. *Clin Cancer Res.* 709
- 2011;17:505–13. 710
44. Trollet J, Hupe P, Huon I, Lebigot I, Decraene C, Delattre O, et al. 711
- Genomic profiling and identification of high-risk uveal melanoma 712
- by array CGH analysis of primary tumors and liver metastases. 713
- Invest Ophthalmol Vis Sci.* 2009;50:2572–80. 714

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